Corrections

Lecithin Retinol Acyltransferase Forms Functional Homodimers, by Wan Jin Jahng, Eric Cheung, and Robert R. Rando*, Volume 41, Number 20, May 21, 2002, pages 6311−6319.

Page 6315. In the legend of Figure 2, all millimolar (mM) concentrations should be micromolar (μM).

Page 6316. In the legend of Figure 4, the concentrations of RPE and BMH in lane 3 should be 10 μM and those of RPE and BMH in lane 4 should be 50 μM.

Page 6317. In footnote c of Table 1, C# is carbamidomethylated cysteine.

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Membrane Orientation and Position of the C2 Domain from cPLA2 by Site-Directed Spin Labeling, by April A. Frazier, Mark A. Wisner, Nathan J. Malmberg, Kenneth G. Victor, Gail E. Fanucci, Eric A. Nalefski, Joseph J. Falke,* and David S. Cafiso,* Volume 41, Number 20, May 21, 2002, pages 6282−6292.

Page 6283. In the introduction to our paper, we indicate that a model generated in a previous EPR study [Ball, A., Nielsen, R., Gelb, M. H., and Robinson, B. H. (1999) *Proc. Natl. Acad. Sci. U.S.A. 96*, 6637−6642] places the cPLA2 C2 domain so that Ca^{2+}-binding loop 3 penetrates into the bilayer, while loop 1 does not. Our statement was based on the model shown in Figure 3 of the earlier work by Ball et al. While our statement accurately reflects the model presented in Figure 3 of Ball et al., which they describe as the optimal fit to their data, more careful examination of their oxygen collision data indicates that it is consistent with the penetration of Ca^{2+}-binding loops 1 and 3 into the bilayer. Thus, in contrast to the comments in the text of our paper and Figure 3 of Ball et al., the different EPR methods utilized in the two studies both indicate that Ca^{2+}-binding loops 1 and 3 penetrate into the bilayer. The two models differ in the orientation of the domain relative to the membrane: in our model the long axis of the domain is tilted further toward the membrane normal relative to the optimal orientation proposed by Ball et al.

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